

# Journal of Experimental Research

SEPTEMBER 2023, Vol 11 No 3

Email: editorinchief.erjournal@gmail.com

editorialsecretary.erjournal@gmail.com Received: August, 2023 Accepted for Publication: September, 2023

## IDENTIFICATION AND PREVALENCE OF *TAENIA* SOLIUM EGGS IN FAECES OF PIGS IN WUKARI, SOUTHERN TARABA, NIGERIA

\*<sup>1</sup>Awujo Nkem Chinedu, <sup>2</sup>Nyenbenso Nancy and <sup>3</sup>Ishaku Fyinbu Dennis

<sup>1,2,3</sup>Tropical Disease Research Laboratory, Department of Microbiology, Federal University Wukari, P.M.B. 1020, Wukari, Taraba State

\*Author for Correspondence: chineduawujo@gmail.com;

#### ABSTRACT

This study was carried out to identify T. solium eggs in feacal droppings of one hundred (100) unrestrained pigs in different sampling areas of WapanNghaku and Mission Quarters in Wukari, Southern Taraba State, Nigeria, using the formol-ether-sedimentation and direct wet mount techniques. Taenia solium (T. solium), also known as pork tapeworm, is a segmented intestinal parasite of humans and pigs that is endemic in many developing countries. The prevalence of eggs in both diagnostic methods was compared using percentages. The prevalence of *T. solium* eggs using the formol-ether-sedimentation and direct wet mount technique was 48% and 32% respectively. In both diagnostic isolation methods, the rate was highest in the WapanNghaku (formol-ethersedimentation: 56%; direct wet mount: 40%) than in the Mission Quarters areas (formol-ethersedimentation: 40%; direct wet mount: 20%). This study which describes for the first time, the study on T. solium in nomadic pigs and techniques to identify their eggs in faeces and determine their prevalences in Wukari, has improved data on the epidemiology of T. solium by showing that nomadic pig farming in Wukari predisposes transmission of taeniasis in the study population of pigs that serve as reservoirs of T. solium eggs. Furthermore, these eggs can be isolated with more accuracy using the formol-ether-sedimentation technique which is simple. Whilst a robust surveillance data is advocated, there is need to adopt a quality intensive system of pig management to maintain good hygienic environment and disease free pork for consumption in the study areas.

Keywords: Identification, sedimentation, wet mount, Taenia solium, faeces, pigs

### **INTRODUCTION**

*T. solium* is a zoonotic pork tapeworm of the phylum plathyhelminthes and class Cestoda (Otubanjo, 2013; Weka et al. 2020). It is cosmopolitan in distribution but endemic in many developing countries of Latin America, Africa and South East Asia (Zammarchi et al. 2013; Okello et al. 2014; Rodriguez-Morales et al. 2018; Alarakol et al. 2021).Nigeria is among the significant pig rearing countries in Africa (Robinson et al. 2014). T. solium taeniasis/cysticercosis/ neurocysticercosis is an important but neglected public health problem and serious socialeconomic obstacle for pig breeders in many African countries including Nigeria (Igbokwe and Maduka, 2018; Melki et al. 2018). The number of live pigs in Nigeria was reported to increase from 8,005 heads in 2011 to 8,092 heads in 2021,

growing at an average annual rate of 4.09% (Sasu, 2023).

Infection with *T. solium* tapeworm (taeniasis) occurs when a person eats raw or undercooked infected pork containing the parasites' larval cysts or cysticerci (Otubanjo, 2013). Within four months, the larvae evaginate in the stomach and duodenum, attach to the intestinal wall and develop into adult tapeworms with gravid segments. Upon maturity, a single tapeworm can shed as many as 1000-2000 eggs per day. Each egg encloses an invasive hexacanth, or onchosphere which along with the distal gravid proglottids, are shed sporadically into the environment in the faeces of the tapeworm carrier. Although many eggs are discharged from the proglottids through an anterior pore, some remain within the uteri and remain viable for many months protected by the environmental factors and faeces and, from where

An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 This work is licenced to the publisher under the Creative Commons Attribution 4.0 International License. 165 habitats and practices, T. solium eggs may also State, Nigeria. Wukari is geographically situated infect humans via ingestion of contaminated food between Latitude 7°53 42 North and Longitude or water. Ingested eggs develop into viable larvae 9º47 59 East. It has an area of 4,308km<sup>2</sup> and a (cysticerci) in tissues causing human population of 241, 546 people made up of ethnic cysticercosis. Autoinfection involves the Jukun, Kuteb, Fulani, Hausa, Shombo and Tiv. retrograde transmission of proglottids from the Prominently, the Jukuns and other tribes in their intestines to the stomach via reverse peristalsis minorities that inhabit Wukari are involved in (Otubanjo, 2013; Zammarchi et al. 2013; WHO, occupations such as agriculture, trading and 2023).

Pig rearing is an important business globally (Robinson et al. 2014). However, cysticercosis Ethical approval and faecal dropping reduces the market value of pigs and makes pork collection unsafe to eat (WHO, 2023). Socioeconomic conditions such as poor environmental hygiene obtained from the Ethical Board of the and management practices are huge contributory risk factors of pig infections with parasites (Kungu et al. 2015; WHO, 2023).

in small holder areas as scavengers of refuse droves were painstakingly and patiently followed dumps (Tidi et al. 2011). In areas where open as they wandered about and care was taken to defecation is still practiced, pigs freely forage human refuse and contaminated water or food. The pigs raised in this way are sold in the local marked thereafter. The faecal dropping of each pig market either directly to butchers or via traders passed out in soil was collected as the drove who travel from one farm to another to purchase roamed the roadsides, backyards, fields and home pigs thereby perpetuating the transmission of T. solium (Tidi et al. 2011; Karshima et al. 2013; Bernard et al. 2015). Investigations have shown Quarters). A total of one hundred (100) droppings varied prevalence of parasitosis in Nigeria. For from 100 pigs were collected. A separate example, Olaniyi (2014) reported a rate greater disposable glove and clean pre-labelled specimen than 17% in pigs in Kwara State, Nigeria while bottle was used for each faecal dropping. The Bernard et al. (2015) recorded a prevalence of sample bottles were well sealed and transported to 32.5% amongst pigs in Pankshin, Plateau State of the laboratory for analysis within 24 hours. Nigeria.

As far as could be ascertained, there is no existing research that has identified the presence and/or prevalence of T. solium eggs in nomadic pigs in Wukari. Pigs are in abundance in Wukari and roam the entire neighborhood where they are raised under poor sanitary conditions for human consumption. Thus, the present study was undertaken to isolate and identify T. solium eggs in faecal droppings of free or unrestricted pigs using the formol ether sedimentation and direct mount techniques.

### **MATERIALS AND METHODS**

#### Study Area:

The study was conducted in Wukari town in

they can act as sources of zoonoses. In unsanitary Wukari Local Government Area (LGA) of Taraba hunting (Oruonye and Abbas, 2011).

Approval for this research project was Department of Microbiology, Federal University Wukari. Twenty-five pigs each from each sampling area were randomly selected and In Nigeria, pigs have been found to be reared monitored during sample collection. Different ensure that droppings were collected immediately they were passed out and the sampled pig ink surroundings of four (4) areas in Wukari town (WapanNghaku, Ken Kisu, East and Mission

#### Laboratory examination of droppings

The faecal droppings were first examined macroscopically for the presence of whole worms or segments. Thereafter, the droppings were processed for microscopy using the direct wet mount (DWM) and formol-ether sedimentation (FES) techniques described below (Cheesbrough, 2006).

To prepare a direct saline wet mount, one drop of physiological saline solution (0.85% w/v sodium chloride) was placed on to a clean and grease-free microscope slide. Thereafter, with the aid of an applicator stick, a small amount of the faecal dropping corresponding to a match stick head was added to it and mixed thoroughly. Finally, a cover slip (22mm by 22mm) was applied over this uniform suspension and microscopically

An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 166 This work is licenced to the publisher under the Creative Commons Attribution 4.0 International License.

detection of T. solium eggs.

In order to isolate and identify *T. solium* eggs x40 objectives. in the faecal droppings, a small quantity of the sample was transferred into a 50 mL beaker containing 10 mL of physiological saline. The solution was thoroughly vortexed with a glass rod and the emulsion filtered through fine mesh gauze test showed that out of one hundred (100) faecal into a 15 mL conical centrifuge tube. The droppings examined for the presence of T. solium suspension was centrifuged at a relative eggs, the prevalence of eggs in droppings was centrifugal force of 600g (2000 revolution per 48% (Table 1). The highest prevalence (56%) was minute, rpm) for 10 minutes to yield about 0.75 from the WapanNghaku droppings while the least mL of sediment. The supernatant was decanted prevalence (40%) was from Mission Quarters and the sediment washed with 10 mL of saline droppings. The result also showed equal prevalent solution. This was centrifuged and washed again rates of *T. solium* eggs (48%) from the Ken Kisu until a clear supernatant was obtained. After the and East area droppings. last wash, the supernatant was decanted and 10 mL of 10% buffered formalin was added to the number of T. solium eggs was found to be highest sediment, mixed and left to stand for 5 minutes for in the droppings collected from the WapanNghaku fixation to occur. Thereafter, 4 mL of diethyl ether area (40%) while the droppings collected from the was added to the sediment and the tube was Mission Quarters contained the least eggs (20%). stoppered, contents vigorously shaken and The prevalence of eggs in Ken Kisu and East centrifuged at 1500 rpm for 10 minutes. droppings was 32% and 36% respectively (Table Resultantly, four layers consisting of top layers of 1). diethyl ether and debris plug, layers of formalin and sediments were obtained. The debris plug was the percentage detection of T. solium eggs was carefully freed from the side of the centrifuge tube highest in the faecal droppings identified using the by ringing with an applicator stick while the top FEC technique and least in those detected with the three layers were decanted. The remaining direct wet mount technique (Table 1). sediment was mixed with a pipette and one drop

examined using the x10 and x40 objectives. The each transferred to a drop of saline and iodine on a goal of this method was to increase the glass slide and mixed. The two drops were covered translucency of the sample and enhance the with cover slips and microscopically examined for the presence of *T. solium* eggs using the x10 and

#### RESULTS

The result of the formol-ether sedimentation

Using the direct wet mount technique, the

Comparatively, in each of the areas sampled,

Sample area	Number examined	Number of positive samples Formol-ether sedimentation Direct wet mount	
WapanNghaku	25	14(56.0)	10(40.0)
Ken Kisu	25	12(48.0)	8(32.0)
East	25	12(48.0)	9(36.0)
Mission	25	10(40.0)	5(20.0)
Quarters			
Total	100	48(48.0)	32(32.0)

Table 1: Identification of *T. solium* eggs in faecal droppings using the formol-ether sedimentation and direct wet mount techniques

Figures in parentheses represent percentages

An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 167 This work is licenced to the publisher under the Creative Commons Attribution 4.0 International License.

#### DISCUSSION

Mature eggs that are shed into the environment might remain active for months under favourable environmental conditions (WHO, 2023). The determination of prevalences of T. solium eggs using the FES diagnostic method was higher than when the DWM technique was used because the number of helminthic eggs is often too low to be observed microscopically in DWM while the use of FES increases the percentage detection of the copro-microscopic technique. This implies that the FES method concentrated the helminth eggs by taking advantage of their high specific gravity compared to water. The natural inclination of these eggs to settle (sediment) in aqueous solutions is accelerated through centrifugation. Formalin fixed the eggs rendering them non-infectious and preserved their morphology while ether was used to extract debris and fat from the faeces thus enhancing parasite identification (Cheesbrough, 2006).

Just as Saelens et al. (2022) observed, several techniques that have been employed by researchers to identify taeniid eggs seldom have standardization, performance evaluation and viability assessment significance and hinder understudy comparisons even as prospective investigators find it difficult to comprehend and decide the best method to use to determine environmental contamination by eggs of Taenia sp. No supporting and appropriate reference article as far as could be ascertained, was found to enable epidemiological studies to be made for FEC better than DWM in detecting T. solium eggs in contaminated soils. Most of published literature usually utilize a single diagnostic procedure to detect taeniid eggs in animal, water, food and soil environments (Adenusi et al. 2015; Bernard et al. 2015; Guggisberg et al. 2020) and where two or more methods have been used, the objectives were not to compare the detection rates based on recovery methods (Satchwell, 1986; Maikai et al. 2012; Jimenez et al. 2016; Aghaindum et al. 2019). It is noteworthy that the determination of the prevalence of T. solium eggs was successfully carried out in this present study using both the FES and DWM diagnostic techniques of isolation.

Generally, the egg positivity in the samples using either of the two diagnostic techniques was high in the study area due to the increased risk of

parasite transmission arising from the large population of freely roaming pigs, their feeding habit and circumstances of raising them under this free range system (Kungu et al. 2015). In the areas they roamed, some households had no toilets and most of the children defecate on near-by shrubby areas or in the bushes, a high risk factor for contamination and transmission (Jansen, 2021). Under these conditions, roaming pigs are likely to feed on faecal materials that might contain *T. solium* eggs. This may be the reason the prevalence of eggs was least in Mission Quarter as the living standards especially toilet facilities and wastes disposal practices were observed to be better than in the other areas.

#### CONCLUSION

It is evident in this study that pigs are important sources of *T. solium* eggs. To reduce the risk of transmission to humans, local breeders are advised to vaccinate and/or de-worm their pigs as core "rapid impact" intervention schemes. Open defecation and dumping of faecal refuse should be discouraged through vigorous community health education as supporting measures. Fundamental societal changes such as thorough cooking of meat before consumption, improved husbandry management and meat inspection practices, are hereby advocated since pork meatisa widely accepted delicacy in Wukari.

## CONFLICT OF INTEREST AND FUNDING STATEMENT

The authors declare no conflict of interest. They also did not receive any funding whatsoever in the course of this research.

#### STATEMENTS AND DECLARATIONS

The authors declare that this work has neither been published before nor under consideration for publication anywhere else. The manuscript has also been read and approved by all authors.

#### REFERENCES

Adenusi AA, Abimbola WA, Adewoga TOS. (2015). Human intestinal helminth contamination in pre-washed, fresh vegetables for sale in major markets in Ogun State, southwest Nigeria. Food Control. 50: 843-849.

Aghaindum AG, Atud AQ, Nadege OA. (2019). Implications of soils around domestic water points

An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 This work is licenced to the publisher under the Creative Commons Attribution 4.0 International License. 168 Yaounde (Cameroon). Journal of Water Health. 17: 318-328.

- Alarakol SP, Bagaya BS, Yagos WO, Odongo-Aginya EI. (2021). Prevalence and risk factor associated with T. solium cysticercosis among pig farmers in two districts (Amuru and Gulu) in Northern Uganda. Journal of Parasitology and Vector Biology. 13(1): 25-34.
- Bernard AN, Daminabo V, Ekam E, Okankwo EC, Nwuzo AC, Afiukwa FN, Agah MV.(2015). Prevalence of intestinal parasites in faecal droppings of swine in Pankshin Urban, Pankshin Local Government Area, Plateau State, Nigeria. American Journal of Life Sciences. 3(2):119-122.
- Cheesbrough M. (2006). Parasitological tests. In: District Laboratory Practice in Tropical Countries (Part 1, 2nded.). Cambridge University Press, Edinburg Building, Cambridge CB2 1IR, United Kingdom. Pp. 191-200
- Guggisberg A, Alvarez RC, Kronenberg P, Miranda N, Deplazes P. (2020). A sensitive, one-way sequential sieving method to isolate helminths' eggs and protozoal oocysts from lettuce for genetic identification. Pathogens. 9: 624.
- Igbokwe IO, Maduka CV. (2018). Disease burden affecting and challenges. Revue D'élevage et de Médicine Vétérinaire des pays Tropicaux. 71(1-2):87-95.
- Jansen F, Dorny P, Gabri"el S, Dermauw V, Johansen MV, Trevisan C. (2021). The survival and dispersal of Saelens G, Robertson L, Gabriel S. (2022). Diagnostic tools Taenia eggs in the environment: what are the implications for transmission? A systematic review. Parasite Vectors. 14:88.
- Jimenez B, Maya C, Velasquez G, Torner F, Arambula E, Barrios JA, Velasco M. (2016). Identification and quantification of pathogenic helminth eggs using a digital image system. Experimental Parasitology. 166:164-172.
- Karshima NS, Bobbo AA, Udokainyang AD, Salihu A. (2013). Taenia solium cysticercosis in pigs slaughtered in Ibi Local Government Area of Taraba State, Nigeria. Journal of Animal Science Advances. 3(3):109-13.
- Kungu JM, Michel O, Ejobi F. (2015). Status of Taenia solium cysticercosis and predisposing factors in developing countries involved in pig farming. International Journal of One Health.1:6-13.
- Maikai BV, Elisha IA, Baba-Onoja E. (2012). Contamination of vegetables sold in markets with Nigeria. Food Control. 28:345-348.
- Melki J, Koffi E, Boka M, Soumahoro M-K, Jambou R. (2018). Taenia solium cysticercosis in West Africa: status update. Parasite. 25:143.

- in the spread of intestinal parasites in the city of Okello A, Ash A, Keokhamphet C, Hobbs E, Khamlome B, Dorny P, Thomas L, Allen J. (2014). Investigating a hyper-endemic focus of *Taenia solium* in Northern Lao PDR. Parasites and Vectors. 7:143.
  - Olaniyi AJ. (2014). Public health implication of gastrointestinal parasites of pigs in Kwara State, Nigeria. Journal of Environmental Research and Management. 5(7): 0125-0127.
  - Oruonye ED, Abbas B. (2011). The Geography of Taraba State Nigeria: Natures Gift to the Nation. Lambert Academic Publishing Company, Germany. ISBN-13-9783846504512.264p
  - Otubanjo OA. (2013). Intestinal cestodes. In: Parasites of Man and Animals. Palmgrove, Lagos, Nigeria. Pp. 368-371.
  - Robinson TP, Wint GRW, Conchedda G, Van Boeekel TP, Erocoli V, Palamara E, Cinardi G. (2014). Mapping the global distribution of livestock. Plos One. 9(5):e96084.
  - Rodriguez-Morales AJ, Yepes-Echeverri MC, Acevedo-Mendoza WF, Marin-Rincon HA, Culquichcon C, Parra-Valencia E, Cardona JA, Flisser A. (2018). Mapping the residual incidence of taeniasis and cysticercosis in Colombia, 2009-2013, using geographical information systems. Implication for public health and travel medicine. Travel Medicine Infectious Disease, 22: 51-7.
- pig production in Nigeria. Review of current issues Satchwell MG. (1986). An adaptation of concentration techniques for the enumeration of parasitic helminth eggs from sewage sludge. Water Research. 20: 813-816.
  - for the detection of taeniid eggs in different environmental matrices: a systematic review. Food and Waterborne Parasitology. 26:e00145.
  - Sasu DD. (2023). Nigeria: stock of live pigs. Statista. https://www.statista.com/statistics/1297910/stock -of-live-pigs-in-nigeria/.Accessed 9th September 2023.
  - Tidi SK, Ella FA, Ella AB. (2011). Prevalence of gastrointestinal parasites of pigs in Jos, Plateau State, Nigeria. Nigeria Journal of Parasitology. 32(1): 37-40
  - Weka R, Luka P, Ogo N, Weka P. (2020). Taenia solium cysticercosis in pigs and human: a review of epidemiological data in West Africa (1990-2019). In: Overview of Echinococcosis. Books on Demand. doi:10.5772/intechopen.89559. Pp.1-20.
  - World Health Organization (2023). Taeniasis/cysticercosis. https://www.who.int/news-room/factsheets/detail/taeniasis/cysticercosis.
- helminth eggs in Zaria metropolis, Kaduna State, Zammarchi L, Strohmever M, Bartalesi F, Bruno E, Munoz J, Buonfrate D.(2013). Epidemiology and management of cysticercosis and Taenia solium taeniasis in Europe, systematic review 1990-2011. PLoSone. 8(7):e69537.

An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 169 This work is licenced to the publisher under the Creative Commons Attribution 4.0 International License.